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Review

Colorectal cancer chemoprevention: biochemical targets and clinical development of promising agents

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Abstract

Colorectal cancer (CRC) remains a cause of significant mortality in developed countries despite extensive knowledge of its epidemiology and molecular basis. Since multiple molecular steps that collectively bring about this disease are known, its chemoprevention is a realistic proposition. Biochemical targets of CRC chemopreventive agents include carcinogen metabolising enzymes, arachidonic acid metabolism, the transcription factor nuclear factor-kappa beta (NF- κ B), enzymes responsible for polyamine metabolism, and events associated with proliferation and apoptosis of preneoplastic cells. Aspirin, celecoxib, calcium and α -difluoromethylornithine are examples of drugs that have undergone clinical testing. Critical evaluation of these trials allows optimisation of methodologies for clinical advancement of novel chemopreventive agents. Cancer patients can be a suitable cohort of subjects for pilot studies of certain new agents. Such studies and larger trials in high-risk healthy individuals require the stringent use of carefully validated 'preneoplastic' biomarkers which are intrinsically related to defined stages of colorectal carcinogenesis and/or to mechanisms of action of the agent under investigation. © 2001 Elsevier Science Ltd. All rights reserved.

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1. Introduction

The concept of chemoprevention is not new. For many decades, chemicals have been used to prevent dental caries, heart attacks and stroke. Similarly, many individuals modify their diets and lifestyles in an attempt to prevent or delay the onset of disease. Although the history of cancer chemoprevention is relatively brief, many studies have been reported and a few efficacious agents have been identified. The most obvious example is tamoxifen, which can now be considered an established agent in the prevention of breast cancer [1]. Furthermore the vitamin A analogues isotretinoin and retinol palmitate have been demonstrated to prevent second primary cancers in patients with malignancies of the lung and head/neck [2,3]. There is little doubt that the identification of novel chemopreventive agents with efficacy against the major human

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cancers could have an enormously beneficial impact on the public health of Western societies.

This review, which is targeted at a readership concerned primarily with treatment rather than prevention of cancer, will initially attempt to define the parameters which determine whether a chemoprevention study will be useful or not. Therewith, biochemical events associated with colorectal carcinogenesis will be described as potential targets for chemopreventive agents. The final aim is to summarise recent clinical chemoprevention trials of novel agents.

Colorectal cancer (CRC) is the second most common malignancy in developed countries, accounting for approximately 20–30 deaths per 100 000 standard population each year [4]. Although recent years have seen marginal improvements in mortality for white US citizens [4], the prognosis remains poor; similar to that described in 1932 by Dukes [5] and in 1954 by Astler and Coller [6]. The 5-year survival rate for a patient presenting with lymph node involvement (Dukes' C) is approximately 39% following surgery [7]. Although 80% of patients undergoing surgical resection have complete macroscopic clearance of their disease, 50%

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suffer recurrence [8]. Moreover, approximately 25% of patients have advanced disease at presentation [4], when palliative management strategies remain the only option.

The epidemiology and molecular biology underlying CRC have been studied more thoroughly and are better understood than for most other neoplasias. Clinical and molecular evidence suggests that there are several somatic pathways to CRC, and that two of these pathways appear to parallel certain well-defined processes described for inherited forms of the disease [9]. The incidence of CRC has increased rapidly in the latter part of the twentieth century in many countries, including Italy, Japan and urban China [10]. Such changes have occurred within one generation and suggest an important role for environmental influences in the aetiology of this disease. Epidemiological studies have also pinpointed environmental factors, especially dietary, which appear to affect individuals' chances of developing CRC [11]. Diets low in vegetables and folate, and high in fat, red meat and alcohol appear to increase relative risk. Lack of exercise and cigarette smoking are also risk factors. However, the relative risk ratio for each individual factor appears small when compared with factors linked to other cancers, such as smoking which increases the risk of lung cancer almost 20-fold. Red meat intake, for example, may at most double the risk for CRC [11]. Prospective cohort studies have confirmed certain links, such as the protective role of high-dose folate [12], but have failed to substantiate others such as the benefits of dietary fibre [13]. Large prospective studies of dietary risk factors are ongoing, such as the European Prospective Investigation of Cancer, in which data from over 400 000 individuals are being evaluated

Intense research efforts are aimed at integration of our knowledge of molecular pathways with that of the epidemiology of CRC. Integration may ultimately allow the generation of 'risk profiles' for individuals, based on knowledge of family history, lifestyle, dietary factors, genetic analysis and risk/preneoplastic biomarkers (see below). Such profiles may play a significant role in

the selection of subjects for clinical chemoprevention trials. Improved genetic screening for inherited disease traits and an increasing number of pilot screening programmes for sporadic cancer in normal populations [15] are likely to identify increasing numbers of individuals with early stages of colon carcinogenesis. Thus the need for intervention strategies in high-risk individuals and those with preneoplastic lesions will undoubtedly increase.

Cancer chemoprevention can be defined as the inhibition, retardation or reversal of carcinogenic processes by chemical means [16]. Most clinical trials focus on the prevention of carcinogenesis at a premalignant stage or an early phase of malignancy, which is termed 'primary' or 'secondary' chemoprevention, depending on whether it involves normal or high-risk individuals. The treatment with chemopreventive agents of patients who have undergone successful therapy of a primary malignancy but are at increased risk of a second malignancy, is referred to as 'tertiary' chemoprevention [17]. It is notable that the original definition of the term chemoprevention also embraces inhibition of growth and delay of progression of cancers [18].

2. Features of clinical trials

Clinical trials of cancer chemotherapeutic agents have traditionally been divided into three phases; phase I studies to assess potential toxicity and pharmacokinetics; phase II studies to assess potential benefit; and phase III studies to compare the efficacy and toxicity of the new treatment with current therapeutic regimes. The clinical evaluation of agents for chemopreventive properties can also be separated into three phases, albeit with objectives which differ significantly from those in chemotherapy trials. Table 1 summarises salient features of the design of contemporary chemoprevention trials. Unique challenges posed by these studies include trial duration, agent safety and number of subjects enrolled. Although the initiation phase of carcinogenesis may last only hours or days, promotion and progression

Table 1						
Basic generalised	design for	chemoprevention	trials (modified t	from [191)

Phase	Number of subjects	Design	Primary endpoint
I	15–30	Small, short-term pilot study in cancer patients with rapid dose escalation if no toxicity observed	1. Pharmacokinetics
		·	2. Preliminary data on preneoplastic biomarkers
IIa	30–60	Small, short-term dose de-escalation study	Sensitivity and response of preneoplastic biomarkers
IIb	500-1500	Larger, medium-term randomised study in high-risk individuals	Response in definitive preneoplastic biomarkers
III	Several thousand	Large, long-term randomised study with low- and high-risk levels	1. Response in definitive preneoplastic biomarkers
			2. Cancer incidence and mortality

of tumours probably take years, if not decades. Potentially efficacious cancer chemopreventive agents are therefore likely to be administered regularly to healthy individuals who will gain no visible benefit for many years. It is, therefore, important that there be a minimal risk of toxicity. Any adverse effect must be carefully considered in the risk/benefit estimation for the subject population chosen. Moreover, the number of individuals in chemoprevention efficacy trials is generally substantially greater than the number of patients in most chemotherapeutic trials in order to allow observations to reach significance levels.

3. Choice of subjects

Thousands of years ago, Hippocrates and Galen cautioned against the treatment of hidden cancers, arguing that treatment more often than not hastened death [20]. The ethical dilemmas of treating healthy or 'at risk' individuals were highlighted recently by two major lung cancer intervention trials, the α -tocopherol, β -carotene study [21], and the β-carotene and retinol efficacy study [22]. The rationale for the studies was the finding that eating fruit and vegetables is associated with consistently elevated levels of β-carotene and a lower incidence of cancer. However, the results of the trials suggested that β-carotene not only failed to protect against lung cancer, in high-risk groups of smokers and/ or workers occupationally exposed to asbestos, it even increased the risk of developing the disease. Several criticisms have been levelled at the study [23], and consideration of these criticisms may be useful in optimising the design of future CRC chemoprevention studies. Especially noteworthy among the criticisms are the lack of a rigorous scientifically based mechanistic rationale for the trial and the absence of adequate pharmacokinetic/dynamic information to decide proper dose and scheduling. The importance of detailed mechanistic studies, as a prelude to clinical evaluation, in order to minimise the occurrence of unexpected adverse effects was highlighted recently by the suggestion that β-carotene boosts the detrimental effects of cigarette smoke carcinogens by upregulating carcinogen-metabolising enzyme activities [24]. Furthermore, it is conceivable that the fruit or vegetable constituent which is responsible for chemopreventive efficacy is not β -carotene, but a different agent such as its isomer α -carotene [23].

It is also clearly desirable to obtain information on pharmacokinetics and potential toxicity in humans before administering chemopreventive agents over long periods of time to healthy volunteers. Some of the agents under consideration in cancer chemoprevention inhibit steps in the late stages of carcinogenesis and have chemotherapeutic activity in clinical trials. This notion is certainly true for tamoxifen in breast cancer prevention and treatment, and there is evidence in preclinical models and case reports to suggest similar activity for putative CRC chemopreventive agents such as curcumin, an active constituent of the spice turmeric [25,26]. Therefore it may be justifiable to conduct small pilot studies of certain chemopreventive agents in patients with cancer, with measurement of radiological/ chemical tumour markers as well as preneoplastic biomarkers (see below) and pharmacokinetics.

4. Preneoplastic biomarkers

Biomarkers are increasingly used in programmes of screening, chemoprevention and chemotherapy, and there is some degree of confusion in the literature regarding their classification. 'Surrogate endpoint biomarkers' (SEBs) have been identified for several different stages in the carcinogenic process, and represent a means of monitoring disease progression without having to wait for true neoplasia and metaplasia to develop. SEBs can be termed 'preneoplastic biomarkers', and should be distinguished from 'risk biomarkers' and 'tumour markers'. In Table 2 these three basic marker types are defined and their uses outlined. Of these three types, preneoplastic biomarkers are the most useful for assessing the efficacy of cancer chemopreventive agents [27,28].

Carcinoma incidence is the ultimate endpoint of cancer chemopreventive intervention. However, if this acts

Table 2 Types of biomarkers (modified from [27])

Type of biomarker	Stage of disease	Description	Application
Risk biomarker	No disease detectable	Genetic prediposition, past medical history, lifestyle factors, exposure	Useful to molecular epidemiologists to assess risk of developing cancer
Preneoplastic biomarker	Initiation, promotion, progression	Biological alterations representing the early and intermediate stages of carcinogenesis, which may show alterations by chemopreventive agents	Identification of carcinogenesis and chemoprevention
Tumour markers	Established neoplasia, adhesion-migration	Elevated levels associated with particular cancers	Diagnosis and treatment

as the only endpoint, clinical trials are beset by practical difficulties, most notably the prolonged time periods necessary to obtain results. For example, the delay before preventive effects are seen may be 9 years for aspirin [29] and as long as 15 years for high-dose folate [12]. Therefore, sensitive and specific biomarkers are required that accurately reflect the development of malignancy, and may be differentially expressed during the multiple stages of carcinogenesis. In Table 3 some preneoplastic biomarkers of potential use in CRC chemoprevention studies are shown. The frequency of measurement of biomarkers depends on factors such as the accessibility of the biological material required and the potential risk involved in making the measurement. In cardiovascular chemoprevention for example, clinical measurement of arterial blood pressure and laboratory measurement of blood cholesterol are well established as valid predictive markers upon which intervention can be based. Fortunately, in the case of CRC the target organ is anatomically accessible with a relatively small risk from biopsy collection. But it could be desirable to correlate preneoplastic biomarker levels in the target organ with those in peripheral blood. An example of such a correlation has been provided recently in a pilot study [31] preceding a chemoprevention study of broccoli supplements [32]. Although glutathione S-transferase (GST) activity of colon mucosa correlated with that of blood lymphocytes in 29 subjects at increased risk of colorectal cancer, neither was raised significantly by the supplements. Apart from colonic biopsies and blood, patient samples that may be collected for preneoplastic biomarker measurement are urine, stool and colonic lavage fluid.

Preneoplastic biomarkers have not only to be identified, but also painstakingly validated. This task involves the judicious use of the most suitable animal models which reflect the multifactorial nature of human colorectal cancer [33]. Furthermore, the relationship between biomarker and risk of cancer has to be estab-

lished in prospective studies in high-risk individuals. Such studies constitute a significant undertaking, but are necessary to bridge the gap between preclinical model systems and clinical trials.

5. Biochemical targets and chemopreventive mechanisms

5.1. Features of colorectal cancer

Most CRC arises within pre-existing adenomatous polyps or adenomas, which are common lesions, particularly in the elderly [34]. However, it is relatively rare for individual adenomas to progress to malignancy, despite the high rate of cell division [35], and it is, therefore, important to identify 'high-risk' polyps. In order to assess this risk, molecular biological criteria are increasingly used in addition to histological ones. A cell accumulates a combination of genetic defects, including activation of oncogenes and inactivation of tumour genes, to undergo full malignant suppressor transformation [36]. This concept, known as the multistep model of carcinogenesis, has been characterised in some detail for CRC (Fig. 1). Although in reality the progression of events may be less linear and less temporal than this simplified scheme suggests, the model has provided the basis for understanding the interaction between genetic predisposition and environmental factors as outlined in several contemporary reviews [8,9]. Irreversible mutations are known to occur in the initiation phase, followed by continued mutations, uncontrolled cellular proliferation and clonal expansion in the promotion phase. Progression involves genotypically altered cells developing the histological changes associated with CRC.

Identification of high-risk polyps should not only be based on histological criteria and assessment of genetic defects, but might also benefit from consideration of cell signalling pathways, downstream from isolated gene

Table 3
Examples of preneoplastic biomarkers identified for colorectal cancer (modified from [30])

Type of biomarker	Variable measured	Biomarker
Pathological	Histology	Adenoma, aberrant crypt foci
Cellular	Proliferation	BrDU, PCNA, Ki67, lectin labelling
	Differentiation	Lectin labelling
	Apoptosis	TUNEL assay
Biochemical	Arachidonate metabolism	Prostaglandins, COX-2, arachidonic acid, lipooxygenase, leucotrienes
	Polyamine metabolism	Polyamines, ornithine decarboxylase
	Detoxification enzymes	GST, DT-diaphorase
Molecular	DNA methylation, DNA adducts	Methyl groups, MTHFR, MDA-DNA adducts
	Cell cycle	Cyclin D1, TGFα
Genetic	Gene/product	K-ras, APC, DCC

BrDU, bromodeoxyuridine; PCNA, proliferating cell nuclear antigen; TUNEL, terminal deoxyribonucleotidyl transferase mediated nick-end labelling; COX-2, cyclooxygenase-2; GST, glutathione S-transferase; MTHFR, methylenetetrahydrofolate reductase; MDA, malondialdehyde; TGF-α, transforming growth factor-α. APC, adenomatous polyposis coli; DCC, deleted in colorectal cancer.

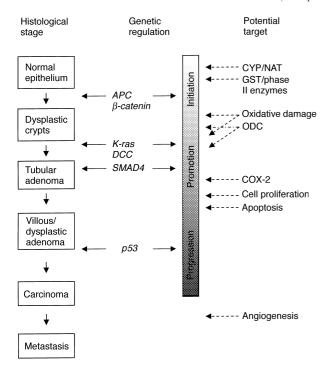


Fig. 1. Multistep model of carcinogenesis with targets for chemopreventive agents (modified from [36]). APC, adenomatous polyposis coli; COX-2, cyclooxygenase-2; CYP, cytochromes P450; *DCC*, deleted in colorectal cancer; GST, glutathione S-transferase; NAT, N-acetyltransferases; ODC, ornithine decarboxylase.

defects which give rise to an abnormal phenotype. It is now recognised that multiple, sequential mutations in genes critical to the control of cellular signal transduction, transcription and thus proliferation and apoptosis are important throughout carcinogenesis [37]. An example is provided by the APC gene which is commonly mutated in polyps and early cancer [38]. Loss of wild-type APC leads to transcriptional activation of cmvc expression through the β -catenin Tcf-4 complex [39], which may affect a number of downstream targets including the initiation factor eIF-4E [40]. Elucidation of the signalling pathway downstream of the wild-type APC gene has pinpointed a crucial role for β -catenin and E-cadherin, the expression of which is frequently abnormal in colorectal cancer [41]. Although APC function may be normal, mutations in β -catenin can result in the abnormalities of adhesion-migration and proliferative signalling found in neoplasia [42].

5.2. Chemopreventive agents

Historically, chemopreventive agents have been categorised as either 'blocking' or 'suppressing' according to the stage of the carcinogenic process with which they interfere [43]. Blocking agents exert their effect at the initiation stage, for example by altering carcinogenmetabolising activities in target tissues or by antioxidant effects. Suppressing agents act at more advanced stages

in the carcinogenic process during promotion and/or progression, by inhibiting cell proliferation and/or inducing apoptosis. Although it is now known that single chemopreventive agents may exert effects at multiple stages (see below), this broad and simple classification remains useful.

It is not yet clear whether highly specific inhibitors, e.g. selective inhibitors of the enzyme cyclooxygenase-2 (COX-2), or agents with more 'pleiotropic' effects, e.g. naturally occurring polyphenols, offer the greatest potential in the chemoprevention of human cancers. In order to effectively counteract processes leading to a disease as multifactorial as cancer, a satisfactory chemopreventive agent may have to possess a variety of mechanistically distinct, but complementary, anticarcinogenic properties. Some important biochemical targets for cancer chemopreventive agents are outlined below.

5.3. Antioxidants and agents which affect xenobiotic metabolism

Reactive oxygen species, such as superoxide anions and hydroxyl radicals, are thought to be involved in carcinogenesis [44]. Consequently, mopping up activated oxygen species is a chemopreventive mechanism displayed by some blocking agents such as flavonoids, vitamin E and isothiocyanates. Phase I drug metabolising enzymes, particularly cytochromes P450, activate many carcinogens, and phase II enzymes, including GST, detoxify carcinogenic metabolites. The balance between carcinogen activation and detoxification is probably a critical arbiter of an individual's risk of developing cancer [45]. This balance is influenced by many blocking agents. One example is indole-3-carbinol which, despite inducing both cytochromes P450 and phase II enzymes, prevents aflatoxin B1-induced hepatocarcinogenesis in rodents [46]. Another example is the isothiocyanate sulphoraphane contained in broccoli, which not only induces xenobiotic conjugating enzymes [47], but also inhibits some cytochrome P450 isoenzymes, prominent among them CYP2E1 [48].

Preclinical and clinical studies have established an association between decreased GST enzyme activity and increased risk for CRC [31]. In subjects at risk of colorectal cancer, the putative chemopreventive agent oltipraz significantly increased GST expression in colon mucosa and in blood lymphocytes [49]. Similarly, administration of non-steroidal anti-inflammatory drugs (NSAIDs) and curcumin has been shown to increase levels of GST isoenzymes in the gastrointestinal tract in preclinical models [50,51].

5.4. Modulators of arachidonic acid metabolism

The arachidonic acid pathway and one of its key enzymes, COX, or prostaglandin H synthase, have

recently received much attention with respect to both the development and prevention of colon cancer. COX catalyses the production of prostaglandins and occurs as two isozymes, COX-1 and COX-2, the latter inducible by infection or inflammation. Several observations support the notion that COX-2 is important in the aetiology of cancer of the colon and several other tissues. A significant increase in COX-2 mRNA has been observed in the majority of colon carcinomas when compared with normal surrounding mucosa, as well as in some adenomas [52,53]. Increased COX-2 expression is often accompanied by elevated levels of eicosanoids including prostaglandins [54,55].

The NSAID, sulindac, which inhibits the expression of COX non-selectively, causes regression of adenomatous polyps in patients with familial adenomatous polyposis (FAP) [56]. Competitive inhibition of COX-2 activity by selective agents such as celecoxib has been shown to suppress the formation of polyps and neoplasias in rodents with carcinogen-induced colon cancer [57,58] and in the minimal intestinal neoplasia (MIN) mouse, a rodent model of the human *APC* gene defect [59]. When APC^{4716} knockout mice which possess a phenotype similar to that of the MIN mouse, were crossed with COX-2 knockout mice, the resultant offspring were significantly protected from tumour development [60].

5.5. Inhibitors of NF-κB activation

Nuclear factor-kappa beta (NF-κB) is a transcription factor pivotal for the expression of many genes regulating proliferation, immunity, inflammatory response and cellular adhesion. It is sequestered in an inactive state in the cytoplasm by an inhibitory protein, IkB [61]. Upon stimulation of the cell by tumour promoters, cytokines or the products of oxidative stress, IkB is phosphorylated by upstream kinases and degraded to release NFκB. NF-κB undergoes translocation to the nucleus where it initiates upregulation of genes containing suitable binding sites. Activation of NF-κB has been shown to inhibit apoptosis [62]. This finding suggests that its inhibition could downregulate genes involved in the promotion and progression of carcinogenesis by restoration of the sensitivity of cells towards apoptotic stimuli [63]. Both aspirin and its hydrolysis product salicylate [64], as well as curcumin [65], interfere with NF-κB activation by inhibiting the IκB kinase (IKK) complex involving a heterodimeric kinase IKK- α and - β .

5.6. Inhibitors of polyamine biosynthesis

Polyamines are short-chain aliphatic molecules required for normal cellular growth. Their concentration in tissues has been shown to correlate with cellular proliferation [66]. Polyamines appear to be involved in

the activation of the proto-oncogenes c-myc and c-fos [67]. Proliferating tissues and tumours differ from normal non-proliferating tissues not only because they produce more polyamines, but also in that they contain individual polyamines at different concentrations. For example, N1-acetylspermidine is generally undetectable in normal mammalian tissues, but is present in high levels in human CRC [68]. The rate-limiting step in polyamine biosynthesis is the enzyme ornithine decarboxylase (ODC), which is constitutively overexpressed in colorectal dysplasia and neoplasia [69,70]. Levels of ODC and polyamines, especially putrescine, are significantly elevated in the colorectal mucosa of individuals with the APC germ line mutation before they develop polyposis [71]. ODC has been suggested to be critical for cell transformation [72], and it might thus act as a preneoplastic biomarker of colorectal carcinogenesis [73]. α-Difluoromethylornithine (DFMO) is an irreversible inhibitor of ODC that is currently under clinical evaluation (see below). Many diet-derived chemopreventive agents, such as genistein, curcumin, indole-3carbinol and green tea polyphenols, have also been shown to inhibit ODC activity [74–77].

5.7. Modulators of cell proliferation and apoptosis

The evidence for antiproliferative and pro-apoptotic properties as important mechanistic determinants of chemopreventive activity is particularly robust in the case of the NSAIDs. Aspirin has been shown to retard proliferation of human colorectal tumour cells by inducing arrest in the G0/G1 phase of the cell cycle and programmed cell death [78]. Salicylate increased the susceptibility of cells at a late stage of neoplastic progression towards induction of apoptosis. Intriguingly, aspirin suppressed the mutator phenotype associated with hereditary non-polyposis CRC by genetic selection for a subset of cells that do not express microsatellite instability [79]. Other chemopreventive agents like curcumin and indole-3 carbinol have also been shown to induce cell cycle arrest and/or apoptosis in cancer cell lines [80,81].

In general, cancer chemopreventive agents have cytostatic rather than cytotoxic properties. A case in point is the specific COX-2 inhibitor SC-58125, which arrested the growth of COX-2-expressing human colon adenocarcinoma cells both *in vitro* and when implanted into nude mice, but it did not affect adenocarcinoma cells lacking COX-2 [82]. This degree of selective activity of a chemopreventive agent is reminiscent of the current emphasis in modern anticancer drug development on compounds which target cellular signalling events selectively [83]. Many of these 'antisignalling molecules', in contrast to their classical anticancer drug precursors, also cause cytostasis rather than cytotoxicity. Such similarities between chemopreventive and antisignalling

chemotherapeutic agents intimate a convergence in the philosophies underpinning new agent development in both areas. This notion is supported by the fact that many cellular signal transduction targets such as *ras*, the mitogen-activated protein kinase pathway and cell cycle regulating molecules, are equally attractive to both areas.

6. Clinical trials

Agents that have entered clinical chemoprevention trials have been chosen on the basis of epidemiological research and activity in preclinical models. The relevance of published clinical trials is discussed in the following paragraphs. Relevant ongoing trials of the same agents, posted on commonly accessed international cancer websites, are shown in Table 4 to allow the reader to monitor future progress.

6.1. Celecoxib

Celecoxib is a highly selective COX-2 inhibitor (see above), which was recently approved in the USA as an adjunct to standard care for patients with FAP based on

the results of a double-blind, placebo-controlled, multicentre trial in 77 patients [84]. Of the two dose levels studied for six months, 400 mg of celecoxib twice daily was found to cause a significant reduction in the number of colorectal polyps on endoscopy. Further NCI-sponsored phase II/III trials in patients with sporadic polyps and hereditary non-polyposis coli are ongoing. Preclinical studies of such agents [82] and clinical experience of NSAIDs such as sulindac [56] would suggest that the chemopreventive effects of highly selective COX-2 inhibitors are likely to be transient, and that polyps may increase in size and number once treatment is curtailed. Long-term safety is being elucidated (see Table 4).

6.2. Aspirin

A number of retrospective epidemiological studies suggest a decreased incidence of colorectal cancer of up to 50% in regular aspirin users [85–87]. One case-control study of 5815 cases of CRC over a 14-year period has suggested that there may be a 9-year delay before any preventive effects are seen from daily aspirin use, and that the reduction in risk may be dose-depen-

Table 4
A selection of clinical trials in progress measuring preneoplastic colorectal biomarkers (obtained from commonly accessed cancer websites, June 2000)

Agent	Trial design ^a	Subject population (recruitment status)	Variables measured ^b	Principal investigator/institution
Celecoxib	Phase I/II	Patients with HNPCC (R)	Pharmacokinetics Histology Others N/D	National Cancer Institute (Bethesda, MD, USA)
Celecoxib	Phase III	Patients with polyps resected (R)	Histology	Bertagnolli M (Boston, MA, USA)
Aspirin	Phase III	Patients with sporadic polyps (R)	Histology	Baron J. (Dartmouth Uni., NH, USA)
DFMO + Sulindac	Phase IIb	Patients with polyps resected previously (N)	Histology Gene/product Apoptosis Arachidonate metabolism Polyamine metabolism	Meyskens F.L. (California Irvine University, CA, USA)
Sulindac versus curcumin/rutin/quercetin	Phase I/II	Normal volunteers (R)	N/D	Shiff S. (Rockefeller University, New York, USA)
Curcumin	Phase I	Patients with advanced CRC (R)	Pharmacokinetics DNA adducts Detoxification enzymes Arachidonate metabolism	Sharma R.A. (Leicester University, UK)
Calcium	Phase III	Patients with sporadic polyps (C)	Histology, others N/D	Baron J. (Dartmouth Uni., NH, USA)
Calcium/Fibre	Phase III	Patients with polyps (R)	Histology, others N/D	Biasco G. (Bologna, Italy)
Folate	Phase II	Patients with polyps resected (R)	Histology, DNA methylation	Mason J. (Boston, MA, USA)

R, recruiting; N/D, not defined; N, not yet recruiting; C, recruitment completed; DFMO, α -difluoromethyl ornithine; HNPCC, hereditary non-polyposis colorectal cancer; CRC, colorectal cancer.

^a See Table 1.

^b See Table 3.

dent [29]. However, the interpretation of retrospective studies is fraught with potential pitfalls, such as individual selection, compliance and confounding variables.

Unfortunately, prospective studies of the incidence of colorectal cancer among users of aspirin or other NSAIDs are still lacking. The only large study published so far has been the Physicians' Health Study [88]. This randomised trial assessed the effects of 325 mg of aspirin taken on alternate days in 22 071 male physicians in the USA. The trial was closed prematurely because of a significant benefit with respect to cardiovascular mortality in the aspirin group. Analysis of 12year follow-up of the aspirin group has not shown any significant effect on CRC incidence [89]. Questions have been raised about the suitability of the subject population, the dose regime adopted, and the fact that the actual period of continual dosing lasted only five years and subsequent data were collected post-trial when follow-up of subjects may have been less stringent.

More recently, strategies to investigate the chemopreventive potential of aspirin have changed. In a pilot phase I de-escalating dose study involving 65 healthy human subjects taking the drug daily for two weeks, preneoplastic biomarkers were measured in colonic tissue in addition to plasma levels of aspirin and salicylic acid [90]. The results of this study suggest that the optimal dose for aspirin as a chemopreventive agent may be 81 mg/day, which is lower than the dosage previously considered necessary for efficacy. Colonic prostaglandin levels continued to be suppressed long after aspirin and salicylic acid had been cleared from the plasma. Subsequent phase II results in patients with colon adenomas have proved encouraging [91], and phase III studies of aspirin and other NSAIDs are ongoing.

6.3. Calcium

Calcium ingested orally forms salts with bile acids and fatty acids once it reaches the colon, and it may thus have a direct anticarcinogenic effect on colonic epithelial cells [92]. Although a few case-control epidemiological studies suggest an inverse association between calcium intake and incidence of colorectal cancer [93], the results are by no means consistent. Similarly, chemoprevention trials have studied histological endpoints in the colon, with mixed results. In certain studies, calcium supplementation decreased epithelial cell hyperproliferation [94,95] and reduced the incidence of metachronous colorectal adenomas [96]. However, two large studies failed to show an effect of calcium on colonic mucosal proliferation [97,98]. Although two recent studies of patients with a previous history of dysplasia or neoplasia have suggested that calcium supplementation may decrease adenoma recurrence [99,100], the optimal dosing regime and the role of confounding dietary constituents remain unclear.

6.4. DFMO

Another agent which has been administered to individuals at risk for CRC is DFMO. Unlike chemoprevention trials in which only histological endpoints were measured, contemporary chemoprevention trials of DFMO have focused on potential toxicity and the tissue levels of polyamines and ODC as preneoplastic biomarkers. In two studies, significant changes in biomarker levels were observed in the target organ, but reversible ototoxicity was experienced at high-dose levels [101,102]. Depending on the subject population being targeted, safety may prove to be the main obstacle in the development of DFMO as a cancer chemopreventive.

6.5. Selenium

Often the secondary analyses of results of negative chemoprevention trials provide surprising results, as exemplified by the selenium skin cancer study [103]. Selenium (200 $\mu g/day$) was administered as a constituent of brewer's yeast to 1312 patients with a history of skin cancers, living in low selenium-intake regions of America. Secondary subset analysis found statistically significant reductions in total cancer mortality, and specifically in the incidence of prostate, lung and colon malignancies.

6.6. Other potential agents

Agents which are currently under consideration for colorectal cancer chemoprevention trials are the dietary substances curcumin, isothiocyanates, vitamin E, vitamin D3, folate, perillyl alcohol and polyphenols from tea [104,105]. On account of their long history of use in food, these compounds are considered harmless, at least at the dose levels consumed in certain diets. Whether this assumption is reasonable for the compounds in their pure forms remains to be demonstrated. Non-dietary compounds under consideration include oestrogens, oltipraz and N-acetylcysteine.

7. Conclusions

With improved molecular and epidemiological knowledge of colorectal carcinogenesis and risk of developing CRC, effective chemoprevention of this common disease is a realistic proposition. Reactive oxygen species, metabolising enzymes, COX-2, NF-κB, polyamines, and key molecules in cell proliferation and apoptosis are potential targets for chemopreventive agents. Integration of knowledge of the molecular biology of CRC with known mechanisms of action of such agents should lead to the discovery of new efficacious

agents and optimisation of clinical trials. CRC chemoprevention trials of aspirin, celecoxib, calcium and DFMO have provided valuable insights into choice of subject cohort, dose levels and the use of biomarkers. The trial of β -carotene in lung cancer patients has highlighted the need for new approaches to subject selection and trial design, especially the desirability of incorporating preclinical and early clinical mechanistic data. There is a need for small pilot studies of putative agents in healthy subjects or cancer patients to obtain information on safety, pharmacokinetics and pharmacodynamics. On account of the prolonged time periods necessary for chemoprevention trials, optimisation will depend on the selection and validation of preneoplastic biomarkers. Adherence to these principles may facilitate the discovery and development of effective chemopreventive agents. The old adage 'to name is to know, a disease known is half cured' may not currently hold for the chemotherapy of CRC. Nevertheless, the optimised understanding of this disease may ultimately contribute to its prevention.

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References

- Fisher B, Costantino JP, Wickerham DL, Redmond CK, Kavanah M, Cronin WM. Tamoxifen for prevention of breast cancer: report of the National Surgical Adjuvant Breast and Bowel Project P-1 Study. *J Natl Cancer Inst* 1998, 90, 1371–1388.
- Hong WK, Lippman SM, Itri LM, Karp DD, Lee JS, Byers RM. Prevention of second primary tumors with isotretinoin in squamous-cell carcinoma of the head and neck. N Engl J Med 1990, 323, 795–801.
- Pastorino U, Infante M, Maioli M, et al. Adjuvant treatment of stage I lung cancer with high-dose vitamin A. J Clin Oncol 1993, 11, 1216–1222.
- Ries LAG, Eisner MP, Kosary CL, et al. SEER Cancer Statistics Review 1973–1997. Bethesda, MD, USA, National Cancer Institute, 2000.
- 5. Dukes CE. The classification of cancer of the rectum. *J Pathol Bacteriol* 1932, **35**, 323–333.
- Astler VB, Coller FA. The prognostic significance of direct extension of carcinoma of the colon and rectum. *Ann Surg* 1954, 192, 846–850.
- Arnaud JP, Buyse M, Nordlinger B. Adjuvant therapy of poor prognosis colon cancer with levamisole: results of an EORTC double-blind randomized clinical trial. *Br J Surg* 1989, 76, 284–289.
- 8. Midgley R, Kerr D. Colorectal cancer. Lancet 1999, 353, 391-
- 9. Potter JD. Colorectal cancer: molecules and populations. *J Natl Cancer Inst* 1999, **91**, 916–932.
- Muir C, Waterhouse J, Mack T, Powell J, Whelan S, Smans M, eds. Cancer Incidence in Five Continents. Lyon, France, International Agency for Research on Cancer, 1987.

- 11. Cummings JH, Bingham SA. Diet and the prevention of cancer. *Br Med J* 1998, **317**, 1636–1640.
- Giovannucci E, Stampfer MJ, Colditz GA, et al. Multivitamin use, folate and colon cancer in women in the Nurses' Health Study. Ann Intern Med 1998, 129, 517–524.
- Fuchs CS, Giovannucci EL, Colditz GA, Willett WC. Dietary fiber and the risk of colorectal cancer and adenoma in women. N Engl J Med 1999, 340, 169–176.
- Riboli E. Nutrition and cancer: background and rationale of EPIC. Ann Oncol 1992, 3, 783–791.
- Kronberg O, Fenger C, Olsen J. Randomised study of screening for colorectal cancer with fecal occult blood test. *Lancet* 1996, 348, 1467–1471.
- Lippman SM, Benner SE, Hong WK. Cancer chemoprevention. J Clin Oncol 1994, 12, 851–873.
- De Flora S, Balansky R, Scatolini L, et al. Adducts to nuclearDNA and mitochondrial DNA as biomarkers in chemoprevention. In Stewart BW, McGregor D, Kleihues P, eds. Principles of Chemoprevention. Lyon, France, IARC Scientific Publication No. 139, 1996, 291–301.
- Sporn MB, Newton DL. Chemoprevention of cancer with retinoids. Fed Proc 1979, 38, 2528–2534.
- Alberts DS. Advanced clinical trial design: the use of surrogate end point biomarkers in cancer chemoprevention trials. In Perry MC, ed. American Society of Clinical Oncology Educational Book. Baltimore, USA, Lippincott, 1999, 76–80.
- Cantor D. Cancer. In Bynum WF, Porter R, eds. Companion Encyclopedia of the History of Medicine Vol. 1. London, UK, Routledge, 1993, 552.
- The Alpha-Tocopherol, Beta Carotene Cancer Prevention Study Group. The effect of vitamin E and beta carotene on the incidence of lung cancer and other cancers in male smokers. N Engl J Med 1994, 330, 1029–1035.
- Omenn GS, Goodman GE, Thornquist MD, Balmes J, Cullen MR, Glass A. Risk factors for lung cancer and for intervention effects in CARET, the beta-carotene and retinol efficacy trial. J Natl Cancer Inst 1996, 88, 1550–1559.
- Knekt P, Järvinen R, Teppo L, Aromaa A, Seppánen R. Role of various carotenoids in lung cancer prevention. *J Natl Cancer Inst* 1999, 91, 182–184.
- Paolini M, Cantelii-Forti G, Perocco P, Pedulli GF, Abdel-Rahman SZ, Legator MS. Co-carcinogenic effect of beta-carotene. *Nature* 1999, 398, 760–761.
- 25. Jiang M-C, Yang-Yen H-F, Yen JJ-Y, Lin J-K. Curcumin induces apoptosis in immortalized NIH 3T3 and malignant cancer cell lines. *Nutr Cancer* 1996, **26**, 111–120.
- 26. Arbiser JL, Klauber N, Rohan R, *et al.* Curcumin is an in vivo inhibitor of angiogenesis. *Mol Medicine* 1998, **4**, 376–383.
- Sharma RA. Cancer chemoprevention: a clinical reality. J Roy Soc Med 2000, in press.
- Einspahr JG, Alberts DA, Gapstur SM, Bostick RM, Emerson SS, Gerner EW. Surrogate end-point biomarkers as measures of colon cancer risk and their use in cancer chemoprevention trials. *Cancer Epid Biomarkers Prev* 1997, 6, 37–48.
- Collet J-P, Sharpe C, Belzile E, Boivin J-F, Hanley J, Abenhaim L.
 Colorectal cancer prevention by non-steroidal anti-inflammatory drugs: effects of dosage and timing. Br J Cancer 1999, 81, 62–68.
- Krishnan K, Ruffin MT, Brenner DE. Clinical models of chemoprevention for colon cancer. *Hematol Oncol Clinics N America* 1998, 12, 1079–1113.
- Szarka CE, Pfeiffer GR, Hum ST, Everley LC, Balshem AM, Moore DF. Glutathione S-transferase activity and glutathione S-transferase μ expression in subjects at risk for colorectal cancer. Cancer Res 1995, 55, 2789–2793.
- 32. Clapper ML, Szarka CE, Pfeiffer GR, *et al.* Preclinical and clinical evaluation of broccoli supplements as inducers of glutathione S-transferase activity. *Clin Cancer Res* 1997, **3**, 25–30.

- 33. Shureiqi I, Reddy P, Brenner DE. Chemoprevention: general perspective. *Crit Rev Oncol/Hematol* 2000, **33**, 157–167.
- Vogelstein B, Fearon ER, Hamilton SR. Genetic alterations during colorectal-tumour development. N Engl J Med 1988, 319, 525–532.
- 35. Bird JP. Observation and quantification of aberrant crypts in the murine colon treated with a colon carcinogen: preliminary findings. *Cancer Lett* 1987, **37**, 147–151.
- 36. Fearon ER, Vogelstein B. A genetic model for colorectal tumorigenesis. *Cell* 1990, **61**, 759–767.
- Loeb L. Cancer cells exhibit a mutator phenotype. Adv Cancer Res 1998, 72, 25–56.
- Powell SM, Zilz N, Beazer-Barclay Y, Bryan TM, Hamilton SR, Thibodeau SN. APC mutations occur early during colectroal tumorigenesis. *Nature* 1992, 359, 235–237.
- 39. He TC, Sparks AB, Rago C, et al. Identification of c-MYC as a target of the APC pathway. Science 1998, 281, 1509–1512.
- Rosenwald IB, Chen J-J, Wang S, Savas L, London IM, Pullman J. Upregulation of protein synthesis initiation factor eIF-4E is an early event during colon carcinogenesis. *Oncogene* 1999, 18, 2507–2517.
- Sparks AB, Morin PJ, Vogelstein B, Kinzler KW. Mutational analysis of the APC/β-catenin/Tcf pathway in colorectal cancer. Cancer Res 1998, 58, 1130–1134.
- Morin PJ, Sparks AB, Korinek V, Barker N, Clevers H, Vogelstein B. Activation of β-catenin-Tcf signaling in colon cancer by mutations in β-catenin or APC. Science 1997, 275, 1787–1790.
- Wattenberg LW. Chemoprevention of cancer. Cancer Res 1985, 45, 1–8.
- 44. Kensler TW, Trush MA, Guyton KZ. Free radicals as targets for cancer chemoprevention: prospects and problems. In Steel VE, Stoner GDM, Boon CW, Kelloff GJ, eds. *Cellular and Molecular Targets for Chemoprevention*. Boca Raton, USA, CRC Press, 1992, 173–191.
- Clapper ML, Szarka CE. Glutathione S-transferasesv biomarkers of cancer risk and chemopreventive response. Chem Biol Interact 1998, 112-112, 377–388.
- Manson MM, Hudson EA, Ball HWL, et al. Chemoprevention of aflatoxin B₁-induced carcinogenesis by indole-3-carbinol in rat liver — predicting the outcome using early biomarkers. Carcinogenesis 1998, 18, 1729–1738.
- Zhang Y, Talalay P, Cho C-G, Posner GH. A major inducer of anticarcinogenic protective enzymes from broccoli: isolation and elucidation of structure. *Proc Natl Acad Sci USA* 1992, 89, 2399–2403.
- 48. Barcelo S, Gardiner JM, Gescher A, Chipman JK. CYP2E1-mediated mechanism of anti-genotoxicity of the broccoli constituent sulforaphane. *Carcinogenesis* 1996, **17**, 277–282.
- O'Dwyer PJ, Szarka CE, Yao K-S, et al. Modulation of gene expression in subjects at risk for colorectal cancer by the chemopreventive dithiolethione oltipraz. J Clin Invest 1996, 98, 1210–1217.
- Van Lieshout EMM, Tiemessen DM, Roelofs HMJ, Peters WHM. Nonsteroidal anti-inflammatory drugs enhance glutathione S-transferase theta levels in rat colon. Biochim Biophys Acta 1998, 1381, 305–311.
- 51. Nijhoff WA, Groen GM, Peters WHM. Induction of rat hepatic and intestinal glutathione S-transferases and glutathione by dietary naturally occurring anticarcinogens. *Int J Oncol* 1993, **3**, 1131–1139.
- 52. Eberhart CE, Coffey RJ, Radhika A, Giardiello FM, Ferrenbach S, DuBois RN. Up-regulation of cyclooxygenase 2 gene expression in human colorectal adenomas and adenocarcinomas. *Gastroenterology* 1994, **107**, 1183–1188.
- Kargmann SL, O'Neill GP, Vickers PJ, Evans JF, Mancini JA, Jothy S. Expression of prostaglandin G/H synthase-1 and -2 protein in human colon cancer. *Cancer Res* 1995, 55, 2556–2559.

- 54. Bennett A, Tacca MD, Stamford IF, Zebro T. Prostaglandins from tumours of human large bowel. *Br J Cancer* 1977, **35**, 881–884.
- Brock TG, McNish RW, Peters-Golden M. Arachidonic acid is preferentially metabolized by cyclooxygenase-2 to prostacyclin and prostaglandin E2. *J Biol Chem* 1999, 274, 11660–11666.
- Giardiello FM, Hamilton SR, Krush AJ, Piantadosi S, Hylind LM, Celano P. Treatment of colonic and rectal adenomas with sulindac in familial adenomatous polyposis. N Engl J Med 1993, 328, 1313–1316.
- Kawamori T, Rao CV, Seibert K, Reddy BS. Chemopreventive activity of celecoxib, a specific cyclooxygenase-2 inhibitor, against colon carcinogenesis. *Cancer Res* 1998, 58, 409–412.
- Reddy BS, Hirose Y, Lubet R, et al. Chemoprevention of colon cancer by specific cyclooxygenase-2 inhibitor, celecoxib, administered during different stages of carcinogenesis. Cancer Res 2000, 60, 293–297.
- Roy HK, Lu K. Nabumetone inhibition of intestinal tumorigenesis in MIN mice: modulation of Bcl-2. Gastroenterology 1997, 112, A647.
- Oshima M, Dinchuk JE, Kargman SL, et al. Suppression of intestinal polyposis in Apc^{Δ716} knockout mice by inhibition of cyclooxygenase 2 (COX-2). Cell 1996, 87, 803–809.
- Simeonidis S, Stauber D, Chen GY, Hendrickson WA, Tahnos D. Mechanisms by which IκB proteins control NF-κB activity. Proc Natl Acad Sci USA 1999, 96, 49–54.
- 62. Beg AA, Baltimore D. An essential role for NF-κB in preventing TNF-α-induced death. *Science* 1996, **274**, 782–784.
- 63. Waddick KG, Uckun FM. Innovative treatment programs against cancer: NF-κB as a molecular target. *Biochem Pharmacol* 1999, **57**, 9–17.
- 64. Yin M-J, Yamamoto Y, Gaynor RB. The anti-inflammatory agents aspirin and salicylate inhibit the activity of IkB kinase-β. *Nature* 1998, 396, 77–80.
- 65. Plummer SM, Holloway KA, Manson MM, et al. Inhibition of cyclooxygenase-2 expression in colon cells by the chemopreventive agent curcumin involves inhibition of NF-kB activation via the NIK/IKK signalling complex. Oncogene 1999, 18, 6013–6020.
- 66. Pegg AE. Recent advances in the biochemistry of polyamines in eukaryotes. *Biochem J* 1988, **234**, 249–262.
- 67. Tabib A, Bachrach A. Activation of the proto-oncogene c-myc and c-fos by c-ras. Involvement of polyamines. *Biochem Biophys Res Commun* 1994, **202**, 720–727.
- Takenoshita S, Matsuzaki S, Nakano G, et al. Selective elevation of the N¹-acetylspermidine level in human colorectal adenocarcinomas. Cancer Res 1984, 44, 845–847.
- Luk GD, Baylin SB. Ornithine decarboxylase as a biologic marker in familial colonic polyposis. N Engl J Med 1984, 311, 80–83.
- Narisawa T, Takahashi M, Niwa M, et al. Increased mucosal ornithine decarboxylase activity in large bowel with multiple tumors, adenocarcinoma and adenoma. Cancer 1989, 63, 1572– 1576.
- Giardiello FM, Hamilton SR, Hylind LM, Yang VW, Tamez P, Casero Jr RA. Ornithine decarboxylase and polyamines in familial adenomatous polyposis. *Cancer Res* 1997, 57, 199–201.
- Auvinen M, Paasinen A, Andersson LC, Hölttä E. Ornithine decarboxylase activity is critical for cell transformation. *Nature* 1992, 360, 355–358.
- Wang W, Liu LQ, Higuchi CM. Mucosal polyamine measurements and colorectal cancer risk. *J Cell Biochem* 1996, 63, 252–257
- Tseng C-P, Verma AK. Inhibition of 12-O-Tetradecanoylphorbol-13-acetate-induced ornithine decarboxylase activity by genistein, a tyrosine kinase inhibitor. *Mol Pharmacol* 1996, 50, 249–257.
- 75. Rao CV, Simi B, Reddy BS. Inhibition by dietary curcumin of azoxymethane-induced ornithine decarboxylase, tyrosine protein kinase, arachidonic acid metabolism and aberrant crypt foci formation in the rat colon. *Carcinogenesis* 1993, 14, 2219–2225.

- Hudson EA, Howells L, Ball HWL, Pfeiifer AMA, Manson MM. Mechanisms of action of indole-3-carbinol as a chemopreventive agent. *Biochem Soc Trans* 1998, 26, S370.
- Gupta S, Ahmad N, Mohan RR, Husain MM, Mukhtar H. Prostate cancer chemoprevention by green tea: *in vitro* and *in vivo* inhibition of testosterone-mediated induction of ornithine decarboxylase. *Cancer Res* 1999, 39, 2115–2120.
- Elder DJ, Hague A, Hicks DJ, Paraskeva C. Differential growth inhibition by the aspirin metabolite salicylate in human colorectal tumor cell lines: enhanced apoptosis in carcinoma and in vitro-transformed adenoma relative to adenoma cell lines. *Can*cer Res 1996, 56, 2273–2276.
- Ruschoff J, Wallinger S, Dietmaier W, Bocker T, Brockhoff G, Hofstadter F. Aspirin suppresses the mutator phenotype associated with hereditary nonpolyposis colorectal cancer by genetic selection. *Proc Natl Acad Sci USA* 1998, 95, 11301–11306.
- Samaha HS, Kelloff GJ, Steele V, Rao CV, Reddy BS. Modulation of apoptosis by sulindac, curcumin, phenylethyl-3-methylcaffeate, and 6-phenylhexyl isothiocyanate: apoptotic index as a biomarker in colon cancer chemoprevention and promotion. *Cancer Res* 1997, 57, 1301–1305.
- 81. Cover CM, Hsieh SJ, Tran SH, et al. Indole-3-carbinol inhibits the expression of cyclin-dependent kinase-6 and induces a G1 cell cycle arrest of human breast cancer cells independent of estrogen receptor signaling. J Biol Chem 1998, 273, 3838–3847.
- Sheng H, Shao J, Kirkland SC, et al. Inhibition of human colon cancer cell growth by selective inhibition of cyclooxygenase-2. J Clin Invest 1998, 273, 2254–2259.
- Gelman KA, Eisenhauer EA, Harris AL, Ratain MJ, Workman P. Anticancer agents targeting signaling molecules and cancer cell environment: challenges for drug development? *J Natl Cancer Inst* 1999, 91, 1281–1287.
- 84. Steinbach G, Lynch PM, Phillips RKS, Wallace MH, Hawk E, Levin B. The effect of celecoxib, a cyclooxygenase-2 inhibitor in familial adenomatous polyposis. *N Engl J Med* 2000, **342**, 1946–1052
- Thun MJ, Namboodiri MM, Heath Jr CW. Aspirin use and reduced risk of fatal colon cancer. N Engl J Med 1991, 325, 1593–1596.
- Giovannucci E, Rimm EB, Stampfer MJ, Colditz GA, Ascherio A, Willett WC. Aspirin use and the risk for colorectal cancer and adenoma in male health professionals. *Ann Intern Med* 1994, 121, 241–246.
- Kune GA, Kune S, Watson LF. Colorectal cancer risk, chronic illnesses, operations and medications: case–control results from the Melbourne Colorectal Cancer Study. *Cancer Res* 1988, 48, 4399–4404.
- 88. Gann PH, Manson JE, Glynn RJ. Low-dose aspirin and the incidence of colorectal tumors in a randomized trial. *J Natl Cancer Inst* 1993, **85**, 1220–1224.
- Sturmer T, Glynn RJ, Lee IM. Aspirin use and colorectal cancer: post-trial follow-up data from the Physicians' Health Study. *Ann Intern Med* 1998, 128, 713–720.

- Krishnan K, Ruffin MT, Brenner DE. Colon cancer chemoprevention: clinical development of aspirin as a chemopreventive agent. J Cell Biochem Suppl 1997, 28/29, 148–158.
- Barnes CJ, Hamby-Mason RL, Hardman WE, et al. Effect of aspirin on prostaglandin E2 formation and transforming growth factor alpha expression in human rectal mucosa from individuals with a history of adenomatous polyps of the colon. Cancer Epidemiol Biomarkers Prevention 1999, 8, 311–315.
- Scalmati A, Lipkin M, Newmark H. Calcium, vitamin D and colon cancer. Clin Appl Nutr 1992, 2, 67–74.
- 93. Potter JD, Slattery ML, Bostick RM, Gapstur SM. Colon cancer: a review of the epidemiology. *Epidemiol Rev* 1993, **15**, 499–545.
- Wargovich MJ, Isbell G, Shabot M. Calcium supplementation decreases rectal epithelial cell proliferation in subjects with sporadic adenomas. *Gastroenterology* 1992, 103, 92–97.
- Bostick RM, Fosdick L, Wood JR, Grambsch P, Grandits GA, Lillemoe TJ. Calcium and colorectal epithelial cell proliferation in sporadic adenoma patients: a randomized double-blinded, placebo-controlled clinical trial. *J Natl Cancer Inst* 1995, 87, 1307–1315.
- Baron JA, Beach M, Mandel JS, van Stolk RU, Haile RW, Sandler RS. Calcium supplements for the prevention of colorectal adenomas. N Engl J Med 1999, 340, 101–107.
- 97. Gregoire PC, Stern HS, Yeung KS. Effect of calcium supplementation on mucosal cell proliferation in high risk patients for colon cancer. *Gut* 1989, **30**, 376–382.
- 98. Baron JA, Toteson TD, Wargovich MJ. Calcium supplementation and rectal mucosal proliferation: a randomized controlled trial. *J Natl Cancer Inst* 1995, **87**, 1275–1277.
- 99. Whelan RL, Hovath KD, Gleason NR, et al. Vitamin and calcium supplement use difluoromethylornithine on rectal mucosal levels of polyamines in a randomized, double-blinded trial for colon cancer prevention. J Natl Cancer Inst 1998, 90, 1212–1218.
- 100. Baron JA (for the Calcium Polyp Prevention Study Group). Calcium supplements for the prevention of colorectal adenomas. N Engl J Med 1999, 340, 101–107.
- Love RR, Jacoby R, Newton MA, et al. A randomized, placebocontrolled trial of low-dose α-difluoromethylornithine in individuals at risk for colorectal cancer. Cancer Epidemiol Biomarkers Prev 1998, 7, 989–992.
- 102. Meyskens FL, Gerner EW, Emerson S, et al. Effect of α-difluoromethylornithine on rectal mucosal levels of polyamines in a randomized, double-blinded trial for colon cancer prevention. J Natl Cancer Inst 1998, 90, 1212–1218.
- Clark LC, Combs GF Jr, Turnbull BW, Slate EH, Chalker DK, Chow J. Effects of selenium supplementation for cancer prevention in patients with carcinoma of the skin. A randomized controlled trial. *J Am Med Assoc* 1997, 277, 1520.
- Kellof GJ, Sigman CC, Greenwald P. Cancer chemoprevention: progress and promise. Eur J Cancer 1999, 35, 1755–1762.
- Kakizoe T. Asian studies of cancer chemoprevention: latest clinical results. Eur J Cancer 2000, 277, 1303–1309.